

**Research paper****Spectrophotometric and Chromatographic  
Methods for the Estimation of Raloxifene  
Hydrochloride in pure form and pharmaceutical  
preparation**

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**ABSTRACT**

**Aim:** To develop simple, accurate and precise spectrophotometric and chromatographic methods were developed and validated for the estimation of Raloxifene Hydrochloride (RXF) in pure and pharmaceutical dosage forms.

**Study design:** spectrophotometric and chromatographic methods.

**Place of Study:** Pharmaceutical Chemistry Department, Faculty of Pharmacy, Cairo University, Egypt.

**Methodology:** The differential spectrophotometric method was based on the measurement of the absorbance difference ( $\Delta A$ ) at 333.4 nm of alkaline raloxifene hydrochloride solutions in 0.1 N NaOH against its acidic solutions in 0.1N HCl. RP-HPLC was developed using benzophenone as an internal standard, where the mobile phase used was acetonitrile: water (50:50, v/v), delivered at a flow rate of 1.2 ml/min on a stationary phase composed of C<sub>18</sub> column; and the detection was carried out at the  $\lambda_{\max}$  of RXF (289 nm).

**Results:** The recovery percentage for RXF was found to be 100.46  $\pm$ 0.65 and 99.96  $\pm$ 0.83 for the two methods, respectively. The methods were validated as per ICH guidelines regarding accuracy, precision and system suitability.

**Conclusion:** All the results obtained were found to be within the acceptable limits. The methods were successful to estimate RXF in bulk powder and pharmaceutical preparation Evista®.

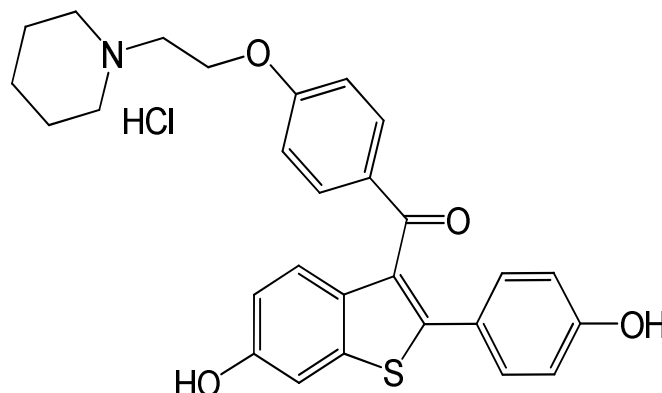
**Keywords:** (Raloxifene Hydrochloride; differential spectrophotometry; absorbance difference; HPLC; benzophenone).

**1. INTRODUCTION**

Raloxifene hydrochloride (RXF) [6-hydroxy-2-(4-hydroxyphenyl)- benzothiophen-3-yl]- [4-[2-(1-piperidyl)ethoxy]phenyl] –methanone hydrochloride] is a selective estrogen receptor modulator acting as an estrogen agonist in bone and in liver it reduces total

22 cholesterol and LDL, but does not increase HDL. It is an estrogen receptor antagonist in  
23 uterine and breast tissues [1]. The chemical structure of RXF is shown in Figure 1. Several  
24 methods have been reported for the analysis of raloxifene hydrochloride in pharmaceutical  
25 dosage form including spectrophotometry [2-6], chromatography [7-11], electrochemistry [12]  
26 and capillary electrophoresis [13].

27 The aim of this work is to develop and validate simple, accurate and precise  
28 spectrophotometric and chromatographic methods were for the estimation of Raloxifene  
29 Hydrochloride (RXF) in pure and pharmaceutical dosage forms.



30

31 **Fig. 1. The chemical structure of Raloxifene hydrochloride (RXF).**

32

## 33 **2. METHODOLOGY**

### 34 **2.1 Chemicals and reagents**

35 Methanol, acetonitrile (Merck) and deionized water were used.

### 36 **2.2 Materials**

37 Raloxifene hydrochloride was kindly supplied by Eli Lilly Co and its purity was  
38 checked by the HPLC method and found to be 99.85±0.81 % [14] and Evista® tablets  
39 labeled to contain raloxifene hydrochloride 60 mg /tablet, Eli Lilly Co.

### 40 **2.3 Apparatus**

41 UV-1650PC, UV - Visible Shimadzu Spectrophotometer (Japan) with matched 1 cm  
42 quartz cells was used for all absorbance measurements. Spectra were automatically  
43 obtained by Shimadzu UV-Probe 2.32 system software. HPLC system, Shimadzu series  
44 (Japan), equipped with PDA detector.

### 45 **2.4 Procedure of the reaction of raloxifene hydrochloride with TCNQ**

#### 46 **2.4.1 Preparation of raloxifene base standard solutions**

47 Twenty mg of pure RXF were accurately weighed, introduced into a 100 ml  
48 volumetric flask and dissolved in methanol to obtain a concentration of (200µg/ml).

49 An accurate weight of benzophenone (200 mg) was introduced into a 100 ml  
50 volumetric flask and dissolved in mixture of acetonitrile: water (50:50, v/v) to obtain a  
51 concentration of (2mg/ml).

#### 52 **2.4.2 Linearity of differential spectrophotometric method**

53 Ten ml aliquot of the standard solution transferred into two 100 ml volumetric flask,  
54 the first was completed to the mark using 0.1 N NaOH and the second one was completed to  
55 the mark with 0.1 N HCl to prepare two working solutions of concentration (20 µg/ml).  
56 Different aliquots of both the alkaline and the acidic solutions (2-10 ml) were transferred into  
57 two separate series of 10 ml volumetric flasks; the volume was completed with 0.1 N NaOH  
58 for the alkaline solution and 0.1 N HCl for the acidic solution. The absorbance difference  
59 ( $\Delta A$ ) of the alkaline solutions of RXF in the sample cell was measured at 333.4 nm relative to  
60 that of the acidic solution of raloxifene hydrochloride in the reference cell. A calibration curve  
61 was plotted, representing the concentration of RXF versus the corresponding absorbance  
62 difference ( $\Delta A$ ), from the average of three experiments.

#### 63 **2.4.3 Chromatographic conditions for RP-HPLC method**

64 The mobile phase used was acetonitrile: water (50:50 v/v). It was filtered through  
65 0.45 µm membrane filter and degassed using ultrasonic path. The system was equilibrated  
66 and saturated with the mobile phase for half an hour before the injection of the samples. The  
67 flow rate was maintained at 1.2 ml/min on a stationary phase composed of C18 column.  
68 Detection was carried out at the  $\lambda_{\max}$  of RXF (289 nm). Benzophenone was used as an  
69 internal standard. System suitability parameters including resolution, number of theoretical  
70 plates, tailing and capacity factors.

#### 71 **2.4.4 Linearity of RP-HPLC method**

72 Different aliquots (2-8 ml) of RXF standard solution (200 µg/ml) equivalent to (400-  
73 1600 µg) were introduced into a series of 10 ml volumetric flasks. Two ml aliquot of  
74 benzophenone solution (internal standard) was added and the volume was completed with  
75 the mobile phase. Twenty micro liter aliquot of each flask was injected into the column. The  
76 previous chromatographic conditions were adopted. The resulting chromatograms, retention  
77 times ( $t_R$ ) of the peaks and the areas under the peaks (AUPs) were recorded. The ratios (R)  
78 of the recorded AUPs of RXF to that of benzophenone were plotted against the  
79 concentration of RXF to obtain the calibration curve from the average of three experiments.

#### 80 **2.4.5 Application to pharmaceutical preparation (Evista® tablets)**

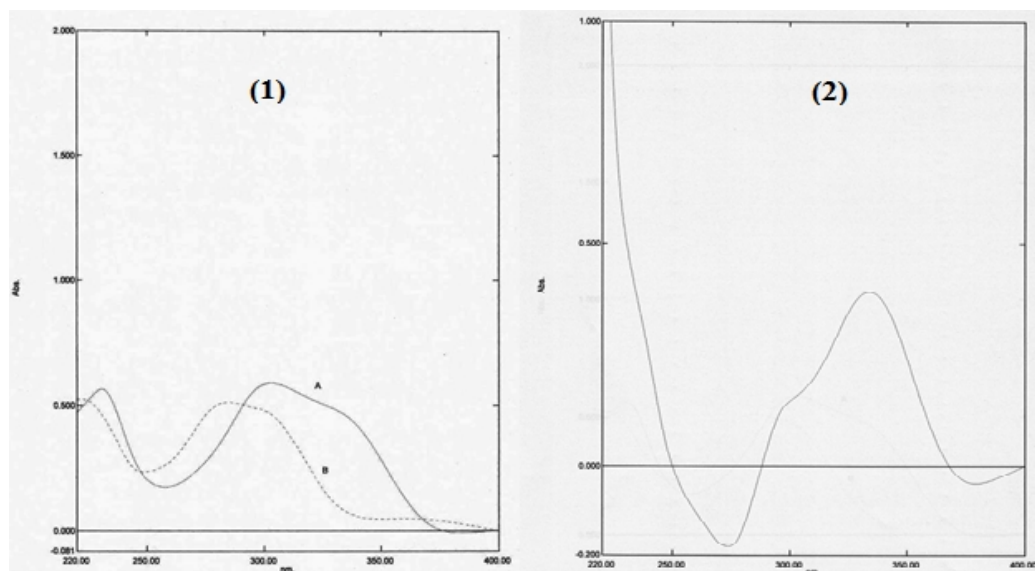
81 An accurate weight of the powdered tablets equivalent to 20 mg of RXF was  
82 transferred into a beaker. Twenty ml aliquot of methanol was added and the beaker was  
83 stirred for 5 minutes with a magnetic stirrer then filtered into a 100 ml volumetric flask and  
84 the volume was completed with methanol to prepare a working solution of (200 µg/ml).  
85 Different aliquots (2- 4 ml) of this solution were transferred into a series of 10 ml volumetric  
86 flasks and completed to the mobile phase. The chromatographic conditions for RP-HPLC  
87 method were applied. The standard addition technique was applied and the regression  
88 equation was used to calculate the recovered concentrations of the labeled and the added  
89 RXF. Further dilution was performed by transferring five ml from the working solution of (200  
90 µg/ml) into two 50 ml volumetric flasks, the first flask was completed to the mark using 0.1 N

91 NaOH and the second one was completed to the mark with 0.1 N HCl to prepare two  
92 solutions of concentration (20  $\mu\text{g/ml}$ ). Different aliquots of both the alkaline and acidic  
93 solutions of RXF equivalent to (60-140  $\mu\text{g}$ ) was transferred into 2 sets of 10 ml volumetric  
94 flasks and completed with the same alkaline or acidic solution. The difference absorbance  
95 ( $\Delta A$ ) of the alkaline solutions of raloxifene hydrochloride in the sample cell was measured at  
96 333.4 nm relative to that of the acidic solution of raloxifene hydrochloride in the reference  
97 cell. The same procedure was repeated using the standard addition technique and the  
98 concentrations of the labeled and added standard RXF could be calculated using the  
99 regression equation.

### 100 3. RESULTS AND DISCUSSION

#### 101 3.1 Differential spectrophotometric method

102 The reported spectrophotometric methods of RXF included colorimetric methods  
103 utilizing different reagents. Those methods are considered to be tedious, time consuming  
104 and of lower reproducibility. So the aim of this work was to develop and validate simple,  
105 accurate and precise spectrophotometric method based the absorbance difference ( $\Delta A$ ) of  
106 UV absorption. The specificity of the difference spectrophotometric determination RXF is due  
107 to its phenolic group which exhibits different spectral changes upon alteration of pH. RXF  
108 exhibits a bathochromic shift of its  $\lambda_{\text{max}}$  at 289 nm in acidic solution to 333.4 nm in alkaline  
109 solution. The difference absorption spectrum ( $\Delta A$ ) of an alkaline RXF solution in 0.1 N NaOH  
110 was relative to an identical concentration in 0.1N HCl in the reference cell shows that the  
111 maximum difference occurs at 333.4 nm, Fig. 2.



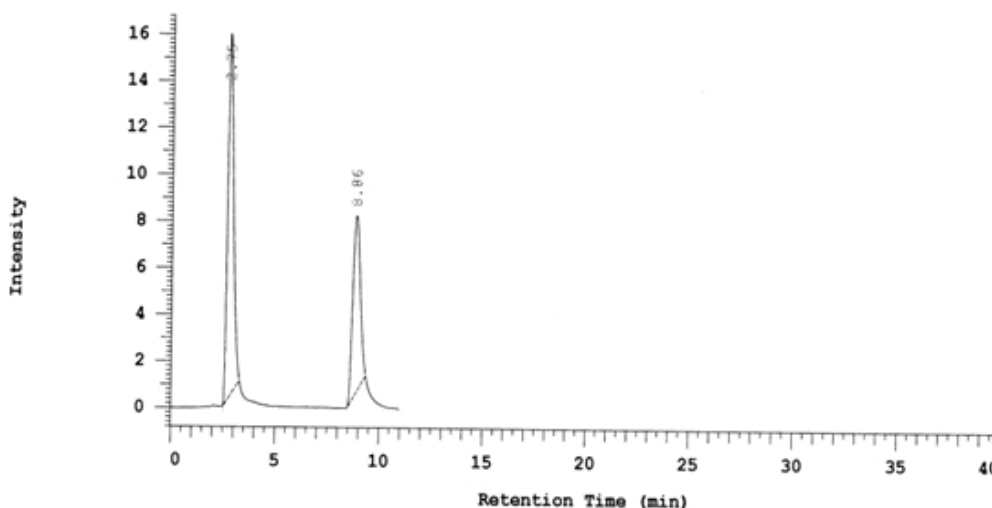
125  
126 **Fig. 2. (1) Absorption spectra of RFX 10 $\mu\text{g/ml}$ , A: in 0.1 N NaOH [—], B: in 0.1 N HCl**  
127 **[...], (2) absorbance difference  $\Delta A$  showing maximum at 333.4 nm.**  
128

#### 129 3.2 RP-HPLC method

130 Several chromatographic methods were reported for the analysis of RXF, but all  
 131 those methods suffered from critical conditions, either by applying gradient elution [8, 11]; or  
 132 by using a mobile phase containing controlled pH buffer [7, 9] which may affect the life time  
 133 of the column. So the aim of this work was to develop and validate accurate and precise  
 134 HPLC method using isocratic elution and a simple mobile phase. In this study, RXF was  
 135 analyzed using a mobile phase composed of (acetonitrile: water, 50: 50 v/v), where it  
 136 showed good resolution between the eluted peaks. Detection was carried at 289 nm. The  
 137 flow rate was 1.2 ml/ min on Waters C18 column.

138 Different binary mixtures of water and methanol were tried, including the (50:50)  
 139 (water: methanol) failed to achieve resolution of the eluted peaks with these systems and the  
 140 detector response was markedly low. Use of water and acetonitrile achieved good  
 141 resolution. Increasing the water ratio decreased the retention times and resulted in peak  
 142 broadening. Different authentic were tried as internal standard but benzophenone was used  
 143 as internal standard with the elution order: raloxifene hydrochloride ( $t_R = 2.75$ ) and  
 144 benzophenone ( $t_R = 8.86$ ). The chromatogram shows complete separation and good  
 145 resolution, Fig. 3.

146



147

148 **Fig. 3. Chromatogram for RP-HPLC showing retention time of RXF ( $t_R = 2.75$ ) and**  
 149 **internal standard benzophenone ( $t_R = 8.86$ ).**

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151 Both methods were applied for the determination of the drug in its pure form and  
 152 pharmaceutical dosage form. The validity of the methods was assessed by applying the  
 153 standard addition technique and the results were shown in Table 1.

154

155 **Table 1. Application of standard addition technique to the analysis of RXF in Evista®**  
 156 **tablets by the proposed methods.**

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Differential spectrophotometric method ( $\Delta A$ )	RP-HPLC method
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Tablet			Standard addition			Tablet			Standard addition		
Claimed*	Found*	Recovery%	Taken*	Found*	Recovery%	Claimed*	Found*	Recovery%	Taken*	Found*	Recovery%
6	6.13	102.17	9.6	9.71	101.15	40	39.87	99.68	60	60.29	100.48
6	6.13	102.17	12	12.01	100.17	60	59.07	98.45	40	39.32	98.30
8	8.21	102.63	7.2	7.23	100.42	60	59.07	98.45	60	60.52	100.87
8	8.21	102.63	12	12.06	100.50	60	59.07	98.45	80	80.49	100.61
10	10.10	101.00	7.2	7.12	98.89	80	78.58	98.23	40	40.39	100.89
14	14.21	101.50	4.8	4.89	101.88	80	78.58	98.23	60	59.36	98.93
<b>Mean</b>		102.02			100.45			98.58			100.01
<b>S.D.</b>		± 0.65			± 0.91			± 0.55			± 1.11
<b>S.E.</b>		± 0.27			± 0.37			± 0.22			± 0.45

158 \* in µg/ml.

159

160 **4. METHOD VALIDATION**

161 Method validation was performed according to ICH guidelines [15] for the proposed  
 162 methods including: linearity, range, accuracy, precision. The results were listed in Table 2.  
 163 System suitability parameters were calculated and compared to USP guidelines [16]. The  
 164 results were listed in Table 3.

165

166 **5. CONCLUSION**

167 The proposed spectrophotometric and chromatographic methods showed several  
 168 advantages over the reported methods. The methods were applied successfully for the  
 169 estimation of raloxifene hydrochloride in pure form and pharmaceutical preparation.

170

171 **COMPETING INTERESTS**

172 Authors have declared that no competing interests exist.

173 **AUTHORS' CONTRIBUTIONS**

174 All the authors have equal share in this manuscript. All authors read and approved the final  
 175 manuscript.

176

177 **Table 2. Assay parameters and validation sheet by applying the proposed methods for**  
 178 **determination of RXF.**

Method	(ΔA) method	RP-HPLC method
<b>Calibration range</b>	4- 20	40- 140

( $\mu\text{g/mL}$ )		
<b>Slope</b>	0.0399	0.0130
<b>Intercept</b>	- 0.0108	- 0.1862
<b>Mean<sup>a</sup></b>	100.15	100.07
<b>SD</b>	$\pm 0.65$	$\pm 0.65$
<b>Accuracy<sup>b</sup></b>	100.46 $\pm$ 0.65	99.96 $\pm$ 0.83
<b>Intra-day precision<sup>c</sup></b>	100.89 / 0.802	100.22 / 0.568
<b>Inter-day precision<sup>c</sup></b>	99.76 / 0.85	100.81 / 0.71
<b><math>r^2</math></b>	0.9993	0.9998

179 <sup>a</sup> Average of three experiments.

180 <sup>b</sup> Mean  $\pm$  SD of five blind concentrations of RXF within concentration range.

181 <sup>c</sup> Mean / RSD of triplicate determination of three concentrations of RXF (n=9).

182

183 **Table 3. System suitability parameters for the RP-HPLC method.**

Parameter	RP-HPLC method for RXF	Reference value [16]
<b><math>t_R</math> (Relative retention time)</b>	2.36	
<b>N (Column efficiency)</b>	2232.89	N > 2000
<b>K' (Capacity factor)</b>	5.00	> 2
<b>T (tailing factor)</b>	1.25	T < 2 T=1 for symmetric peak
<b>Rs (Experimental Resolution)</b>	6.81	Rs >2

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