

1 **A New Anionic Bovine Tendon as Scaffold for the**  
2 **Repair of Bone Defects: A Morphological,**  
3 **Histomorphometric and Immunohistochemical**  
4 **Study**

5  
6 **Rogério L. Buchaim<sup>1\*</sup>, Marcelie P. de O. Rosso<sup>1</sup>, Jesus C. Andreo<sup>1</sup>, Daniela**  
7 **V. Buchaim<sup>2</sup>, Roberta Okamoto<sup>3</sup>, Antonio de C. Rodrigues<sup>1</sup>, Andre L.**  
8 **Shinohara<sup>1</sup>, Jose S. Roque<sup>4</sup>, Domingos D. Roque<sup>2</sup>, Geraldo M. Rosa**  
9 **Junior<sup>5</sup> and Gilberto Goissis<sup>6</sup>**

10  
11 *<sup>1</sup> Department of Biological Sciences (Anatomy), Bauru School of Dentistry, University of São*  
12 *Paulo, Al. Dr. Octávio Pinheiro Brisola 9-75, 17012-901, Bauru, SP, Brazil.*

13 *<sup>2</sup> Medical School, Discipline of Human Morphophysiology, University of Marília (UNIMAR),*  
14 *Marília, São Paulo, Brazil, R. Hygino Muzy Filho, 17525-902, Marília, SP, Brazil.*

15 *<sup>3</sup> Department of Basic Sciences (Anatomy), Araçatuba School of Dentistry, São Paulo State*  
16 *University (UNESP), R. José Bonifácio 1193, 16015-050, Araçatuba, SP, Brazil.*

17 *<sup>4</sup> Anatomy, Northern Paraná State University (UENP), Av. Manoel Ribas 215, 86400-970,*  
18 *Jacarezinho, PR, Brazil.*

19 *<sup>5</sup> Department of Health Sciences, University of Sacred Heart, R. Irmã Arminda 10-50, 17011-*  
20 *160, Bauru, SP, Brazil.*

21 *<sup>6</sup> Faculty of Medicine of São José do Rio Preto (FAMERP), Av. Brigadeiro Faria Lima 5416,*  
22 *15090-000, São José do Rio Preto, SP, Brazil.*

23  
24  
25 *\* Corresponding author: Rogério Leone Buchaim*

26 *E-mail address: rogerio@fob.usp.br*

27 *Phone:*  
28

29 **ABSTRACT:**

30 **Aim:** The process of bone repair is of evident importance in both the clinical and functional  
31 spheres. For this reason, the field of bioengineering has taken it as an object of study, seeking  
32 to perfect the implantation of materials that allow for adequate bone neoformation. This study  
33 investigated the process of bone repair after anionic bovine tendon grafting in rat tibias by  
34 conducting a morphological, histomorphometric and immunohistochemical analysisstudy.

35 **Methodology:** The experimental model consisted of 36 rats randomly divided into two groups: a  
36 control group (CG,  $n=18$ ), in which a surgical cavity in the tibia was filled with blood clots; and  
37 an experimental group (EG,  $n=18$ ), in which a surgical cavity in the tibia was filled with an  
38 anionic bovine tendon graft. In the experimental group, the major axis of the collagen fiber  
39 bundle was placed perpendicularly to the long axis of the tibia. Microscopic, morphometric and  
40 immunohistochemical evaluations were conducted at 7, 15, and 30 days postoperative.

41 **Results:** The analyzes showed an increase and demonstrated increasing bone in bone  
42 neoformation in the experimental group during the assessed periods. There was a significant  
43 difference between day 7 and day 30 and evident vascular proliferation was detected by the  
44 immunohistochemical analysis.

45 **Conclusion:** In conclusion, it can be concluded that the anionic bovine tendon collagen proved  
46 to be an adequate and biocompatible material for efficient bone regeneration, with osteogenic  
47 capabilities that allow it to be used as a scaffold for bone repair.

48

49 **Key-~~W~~ords:** Bone regeneration; ~~B~~bone transplantation; ~~c~~Collagen; ~~i~~mmunohistochemistry;  
50 ~~t~~Tendons; ~~t~~Tissue engineering; ~~t~~Tissue scaffolds.

51

## 52 **1.4 INTRODUCTION**

53

54 Bone defects cause numerous complications that are particularly relevant to the fields of  
55 rehabilitation and orthopedics. Such defects are related to pathological processes and traumatic  
56 or physiological processes, such as fractures, infections, chronic inflammatory diseases,  
57 reduced lean body mass, advanced age, immobility and the effect of glucocorticoid treatments  
58 ~~[[1-4][4-4]~~.

59

60 Bone regeneration in large skeletal defects is a special challenge, as it is essential for adequate  
61 bone repair ~~[5][5]~~ and involves socioeconomic concerns regarding the correct treatment of such  
62 patients ~~[6][6]~~. Physiological bone remodeling is a coordinated process essential to bone repair  
63 and mineral homeostasis, occurring independently at several different anatomical locations ~~[7]~~.

64 | ~~8]~~ [7, 8]. Imbalances in the quantity of removed bone in comparison to newly deposited bone  
65 | lead to reduced amounts of total bone and increased risk of fractures ~~[5]~~ [5].

66 | —

67 | Treatments for such defects require procedures such as autologous or autogenous bone  
68 | grafting, or alternative metal and ceramic grafting, aimed at bone healing and repair ~~[6]~~ [6].

69 | Autogenous grafts have long been considered the gold standard; however, adverse effects in  
70 | the donor site have been observed, leading to the development of biocompatible substitutes for  
71 | this type of graft ~~[9]~~ [9].

72 |

73 | Efforts in this area have focused on tissue engineering and biomaterials ~~[40]~~ in order to study  
74 | the combination of biomaterials and biological systems. The use of devices that reestablish or  
75 | modify tissues or organ function leads to interactions between tissue components and  
76 | biomaterials. This process is associated with the liberation of growth factors in the implantation  
77 | site such as, for example, bone morphogenetic protein-2 (BMP-2), a growth factor that induces  
78 | osteoblast differentiation and promotes bone regeneration ~~[49]~~ [10].

79 | —

80 | Collagen has received special attention from the field of tissue engineering, as it is the most  
81 | abundant protein in mammals, making up to 30% of the protein in the body ~~[11]~~ [44]. It is  
82 | biocompatible and biodegradable, and has low antigenicity and high resistance to traction.  
83 | However, the pure form still presents limitations and its physical and chemical characteristics  
84 | need to be perfected ~~[12,13]~~ [42,43].

85 |

86 | Anionic collagen is created by alkaline treatment, which gives it enhanced piezoelectric  
87 | properties, which is a load change that attracts the osteoblastic action, increasing bone mineral  
88 | density, favoring the deposition of minerals in the organic portion is under pressure, and this  
89 | has widen therapeutic possibilities to bony tissue. a result of selective hydrolysis of  
90 | carboxamide groups of asparagine and glutamine residues from carboxylic collagen ~~[14]~~ [44].

91 | Anionic collagen is capable of ~~guiding~~ attract phosphate and calcium salt deposits in  
92 | accordance with its microfibrillar structure ~~[15]~~ [45]. Glutaraldehyde can ~~also~~ be used ~~as an~~  
93 | ~~alternative~~ in the preparing-ation of the collagenous material, emphasizing its applicability as a

94 biomaterial, since it functions as a stabilizer, reduces immunogenicity and increases resistance  
95 to enzymatic degradation [16-19][16-19]. Used as scaffold systems inserted in bone defects,  
96 such biomaterials are biocompatible and can induce the formation of new bone tissue [20].[20].  
97 \_\_\_\_\_  
98 The biomechanical properties of tendons characterize them as resistant and cable-like, in that  
99 they are formed by dense connective tissue composed of abundant extracellular matrices [21]  
100 [24]. Fiber organization and orientation interfere in bone neoformation [20]-[20]. Thus, it is  
101 essential to study the orientation of collagenous fibers in organic tissue (in the present case,  
102 bone) in order to ensure correct morphological and functional restructuring.

103

104 There are few studies on the behavior of collagen implant tissue derived from bovine tendons  
105 [11][44] and bovine collagen in the form of membranes [22][22], and few descriptions of such  
106 techniques are available. Therefore, the objective of the current study was to analyze the  
107 process of bone repair after anionic bovine tendon grafting in rat tibia, using morphological,  
108 histomorphometric and immunohistochemical analysisstudy.

109

## 110 **2. 2-MATERIALS AND METHODS**

111

### 112 **2.1 2.1-Experimental mModel**

113

114

115 The study was approved by the ethics committee of the University of Marília (Marília, São  
116 Paulo, Brazil). Surgical defects were created in the tibias of 36 male rats (*Rattus norvegicus*,  
117 Wistar), all 60 days old and weighing an average of 245.3 grams.

118

119 The rats were randomly divided into two groups with 18 animals each: a control group (CG), in  
120 which the surgically created cavity was filled with blood clots; and an experimental group (EG),  
121 in which the medullary cavity received an anionic collagen matrix implant made from bovine  
122 calcaneous tendon. During the postoperative period, the rats were kept in individual cages and

123 received *ad libitum* access to food and water. Counting from the day of surgery, six animals  
124 from each group were euthanized by anesthetic overdose at 7, 15 and 30 days postoperative.

125

### 126 **2.2 2-2 Preparation of bBiomaterial**

127

128 This study used fresh bovine tendons (common calcaneous tendons) acquired from a  
129 commercial establishment. The material was prepared and provided by the Chemical Institute of  
130 São Carlos (University of São Paulo, São Paulo, Brazil) in accordance with the literature [16,  
131 23-25][16, 23-25].

132

133 Samples were devitalized by undergoing alkaline sulfate and chlorate solution treatment for 24  
134 hours (to remove cells). The material was neutralized and stabilized in a phosphate buffer, in  
135 accordance with collagen preparation techniques described in the literature by Bet et al. [25]  
136 [25], through selective hydrolysis of asparagine and glutamine amides for 24 hours. Next, the  
137 collagen was balanced with a phosphate buffer, frozen in liquid nitrogen and freeze-dried in an  
138 Edwards Modulyo freeze dryer (Thermo Electron Corporation, Waltham, USA), as described in  
139 a previous study [14][14].

140

141 Differential exploratory calorimetric tests, and transmission electron microscopy (TEM) and  
142 scanning transmission-electron microscopy (SEM) analyses were also carried out, as described  
143 by Bet et al. [25]-[25]. Anionic tendons were then sterilized in ethylene oxide and hydrated  
144 during implantation with a saline solution.

145

### 146 **2.3 2-3 Surgical pProcedures**

147

148 The animals received general anesthesia via intramuscular injections of ketamine hydrochloride  
149 (75 mg/kg; Ceva Santé Animale, Paulínia, Brazil) associated with xylazine hydrochloride (1.5  
150 ml/kg; Ceva Santé Animale, Paulínia, Brazil). A 20 mm longitudinal incision was made in the left  
151 hindlimb, followed by divulsion of muscle tissue surrounding the proximal tibial epiphysis and  
152 separation of the periosteum.

153

154 A bone defect approximately 2.2 mm in diameter was created using a carbide spherical no. 6  
155 steel drill (KG Sorensen, Cotia, Brazil) with a low-speed micromotor (KaVo Dental GmbH,  
156 Biberach, Germany), deeply affecting the medullary cavity. Throughout the procedure, the  
157 surgical site was irrigated with a sterile sodium chloride solution. In the experimental group, the  
158 major axis of the anionic collagen fiber bundle was placed perpendicularly to the long axis of the  
159 tibia (2 mm in diameter), into the defect. In the control group, the defect was maintained with no  
160 biomaterial, and filled only with blood clots. The tissues were reapproximated (including  
161 periosteum) and sutured in layers (Ethicon, Johnson & Johnson Brazil, São José dos Campos,  
162 Brazil).

163

#### 164 **2.4 2.4-Histological Processing**

165

166 Following euthanasia, a portion of the defective tibia was removed, fixated in a 10% formalin  
167 solution for 24 hours, cleansed, and decalcified in a 20% ethylenediaminetetraacetic acid  
168 (EDTA) solution (Merck KGaA, Darmstadt, Germany) for 5 weeks. The solution was changed  
169 once a week, as described in the literature [20][20].

170

171 Next, samples underwent routine laboratory processing and were fixated in paraffin blocks.  
172 Blocks were cut into 6 µm longitudinal sections with a Leica RM 2245 microtome (Leica  
173 Microsystems, Wetzlar, Germany). The samples were then stained with hematoxylin and eosin  
174 and histomorphological analyses were performed using an Olympus BX50 optical microscope  
175 (Olympus Corporation, Tokyo, Japan) [16][16].

176

177 Microscopic analyses were conducted for each group at each postoperative time. This analysis  
178 investigated the superficial surgical site, the lateral edges of residual cortical tissue, the  
179 medullary area adjacent to the superficial surgical site, and the implanted material.

180

#### 181 **2.5 2.5-Immunohistochemistry**

182

183 | The immunohistochemical analysis carried out in the experimental group used two slides from  
184 | each animal to detect immunoperoxidase reactions to identify primary antibodies from the  
185 | following proteins: osteocalcin (OC, sc 240750), vascular endothelial growth factor (VEGF, sc  
186 | 1836), alkaline phosphatase (ALP, sc 79839), tartrate-resistant acid phosphatase (TRAP, sc  
187 | 30832), receptor activator of nuclear factor kB ligand (RANKL, sc 7627) and osteoprotegerin  
188 | (OPG, sc 21038) (Santa Cruz Biotechnology, Santa Cruz, CA, USA). To reveal reactions,  
189 | diaminobenzidine (DAB) was used (Sigma Aldrich, St Louis, MO, USA). Images were recorded  
190 | with an Olympus BX50 microscope (Olympus Corporation, Tokyo, Japan) and photographs  
191 | were taken with an attached digital camera (Olympus DP 71, Tokyo, Japan) with 40x and 100x  
192 | objectives.

193 | Scores of 0 to 3, with 0 = absence of immunostaining (complete absence of immunoreactive  
194 | cells), 1 = low immunostaining (staining in the extracellular matrix and in approximately 1/4 of  
195 | immunoreactive cells), 2 = moderate immunostaining (staining in the extracellular matrix and in  
196 | approximately 1/2 of immunoreactive cells), and 3 = high immunostaining (strong staining in the  
197 | extracellular matrix and in approximately 3/4 of immunoreactive cells).

## 200 | **2.6 2.6-Histomorphometric Aassessment**

201 |  
202 | Quantitative analyses were performed with Image Pro-Plus 6.0 (Media Cybernetics, Silver  
203 | Spring, MD, USA) software. For morphometry, the cortical region where the tibia was perforated  
204 | and the medullary region adjacent to the contralateral intact cortex were analyzed by measuring  
205 | the amount of new connective and bone tissue in the region. The data were subjected to two-  
206 | way analysis of variance (ANOVA) followed by Tukey's test. A significance level was  
207 | established at  $p < 0.05$  for all analyses. The amount of bone tissue and connective tissue  
208 | formed was measured using a light microscope with a 100-point quadrilateral grid system  
209 | coupled with an ocular micrometer, according to the Delesse principle mentioned by previous  
210 | studies [26][26].

## 212 | **3.3 RESULTS**

213

### 214 **3.1.3.1 Histomorphological Analysis**

215

#### 216 **3.1.1 Control Group:**

217

218 Fibrous tissue was present at the superficial surgical site at 7 and 15 days, but was less  
219 prevalent on day 30. Enhanced bone neoformation around the fibrotic area was observed at 7  
220 days and was less evident at 15 and 30 days.

221

222 The medullary cavity was infiltrated by connective tissue. However, there was no bone  
223 differentiation delimitating the blood clot area, still present on day 7. At 15 days, vascular  
224 congestion was prevalent within the trabecular bone and there was rudimentary neocortical  
225 bone with primary bone tissue, which on day 30 was thicker, more organized and mature, with  
226 no well-defined periosteum. Analyses revealed the presence of mononuclear inflammatory cell  
227 infiltrates in neoformed bone at 7 days, which increased in intensity throughout subsequent  
228 days (Fig. 1).

229

#### 230 **3.1.2 Experimental Group:**

231

232 Superficial fibrous tissue was present during the first 15 days, notably more subtle than that in  
233 the control group. Neocortical formation was observed at 15 postoperative days and bone  
234 maturation at 30 days.

235

236 Starting on day 7, tendon fibers were multidirectional and there were fewer perpendicular fibers  
237 in relation to the long axis of the tibia when compared to the time of surgical procedure. Bone  
238 neoformation between anionic tendon fibers was observed on day 15 and was more prevalent  
239 on day 30.

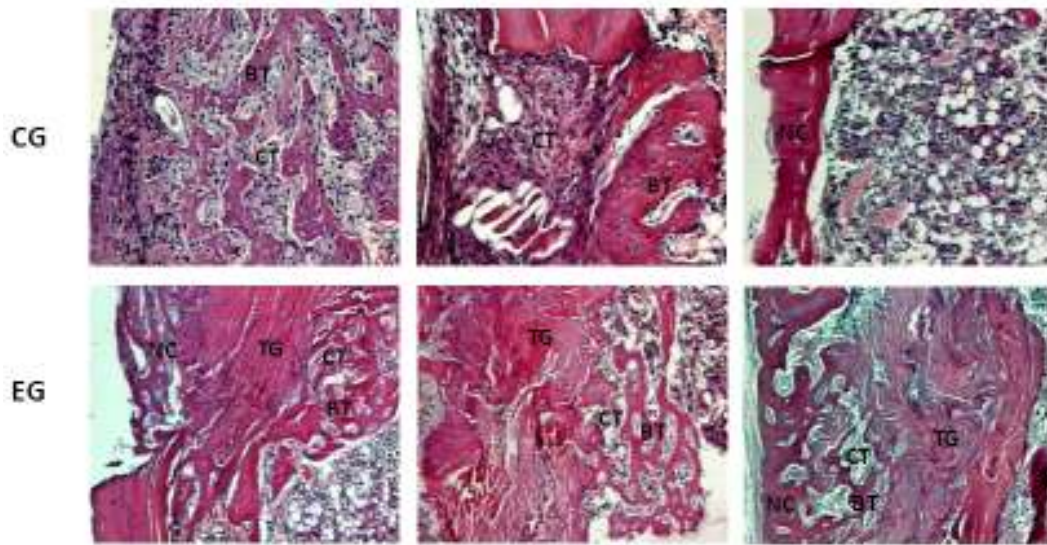
240

241 Increasing bone neoformation and moderate vascular proliferation were observed in proportion  
242 to postoperative time. Osteoblast and osteoclast concentrations were found in the distal end of



243 the tendon. There was accentuated presence of mononuclear inflammatory infiltrates and  
244 interstitial fibrosis on all three studied days (Fig. 1).

245



246

247

248 **FIG. 1.** Histological photomicrography stained with hematoxylin and eosin of control and  
249 experimental groups. 7 days (left), 15 days (middle) and 30 days (right); BT (bone tissue), CT  
250 (connective tissue), NC (new cortical), TG (tendon graft).

251

### 252 **3.2.2 Histomorphometric Analysis**

253

#### 254 **3.2.1 Control Group:**

255

256 Measurement of neoformed bone tissue revealed significant differences between day 30 and  
257 the other assessed periods. Regarding connective tissue, a significant difference was observed  
258 between day 7 and the other investigated periods (Fig. 2).

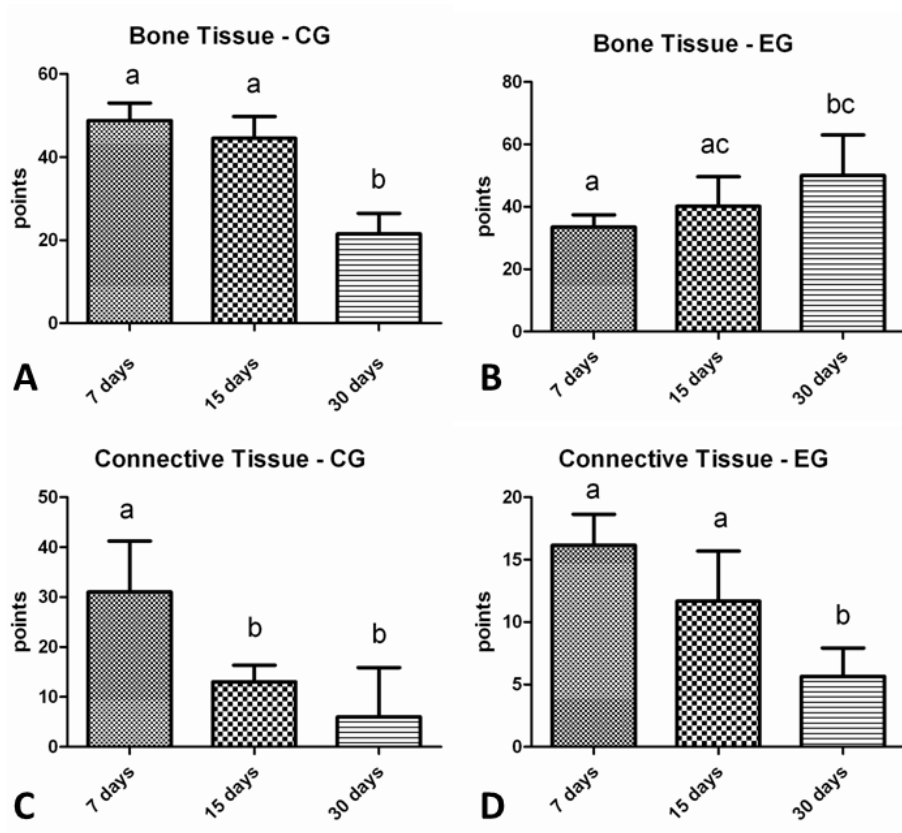
259

#### 260 **3.2.2 Experimental Group:**

261

262 Measurement of neoformed bone tissue revealed a significant difference between day 7 and  
263 day 30. A significant difference was observed regarding connective tissue at 30 days in  
264 comparison to the other assessed periods (Fig. 2).

265



266

267

268 **FIG. 2.** Histomorphometry of the amount of newly formed bone and connective tissue, in  
269 both control (CG (control group)) and EG (experimental groups), at 7, 15 and 30 days  
270 postoperative. Different lowercase letters indicate significant differences among the groups by  
271 means of ANOVA, followed by Tukey's test ( $P < 0.05$ ).

272

### 273 3.3.3 Immunohistochemical Analysis

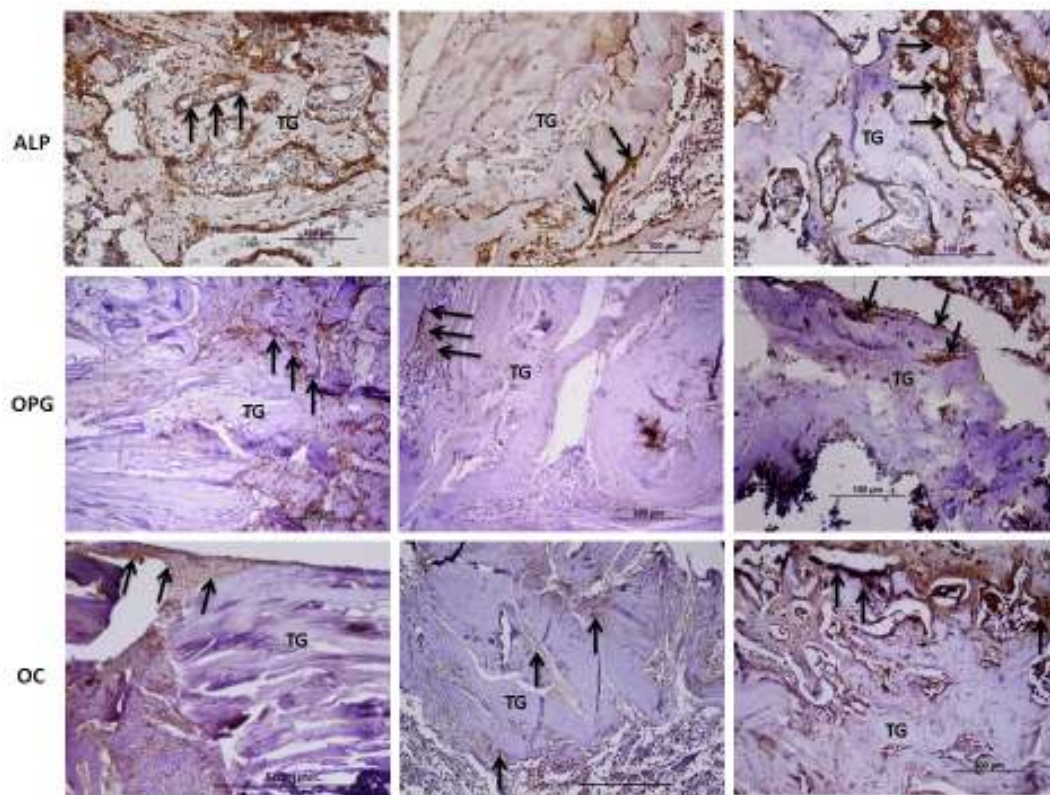
274

275 Immunohistochemical samples for each protein marker used in this study are illustrated in Fig. 3  
276 and Fig. 4. Involved in the osteoclastogenesis regulating mechanism, OPG and RANKL proteins  
277 showed similar levels during all periods; OPG presented peak intensity (score 3) at the

278 | biomaterial-tissue interface at 7 days, and RANKL at 15 days (score 3). Our findings indicated  
279 | that TRAP, a specific protein marker expressing bone reabsorption, was more intensely  
280 | prevalent at 15 and 30 days (score 3). Osteocalcin, a protein from the synthesized extracellular  
281 | matrix secreted during osteoblast differentiation and primarily expressed in the final phase of  
282 | bone formation, was present during all assessed periods, reaching peak intensity at 30 days  
283 | (score 3). In addition to OC, ALP is also used as a marker for osteoblasts with a role in bone  
284 | matrix mineralization, and it was detected during all of the studied periods in expressive  
285 | quantities (score 3). The VEGF protein, a factor expressed by osteoblasts and intimately  
286 | connected with angiogenic processes, was present in all periods, with peak intensity at 15 and  
287 | 30 days postoperative (score 3).

288 | \_\_\_\_\_

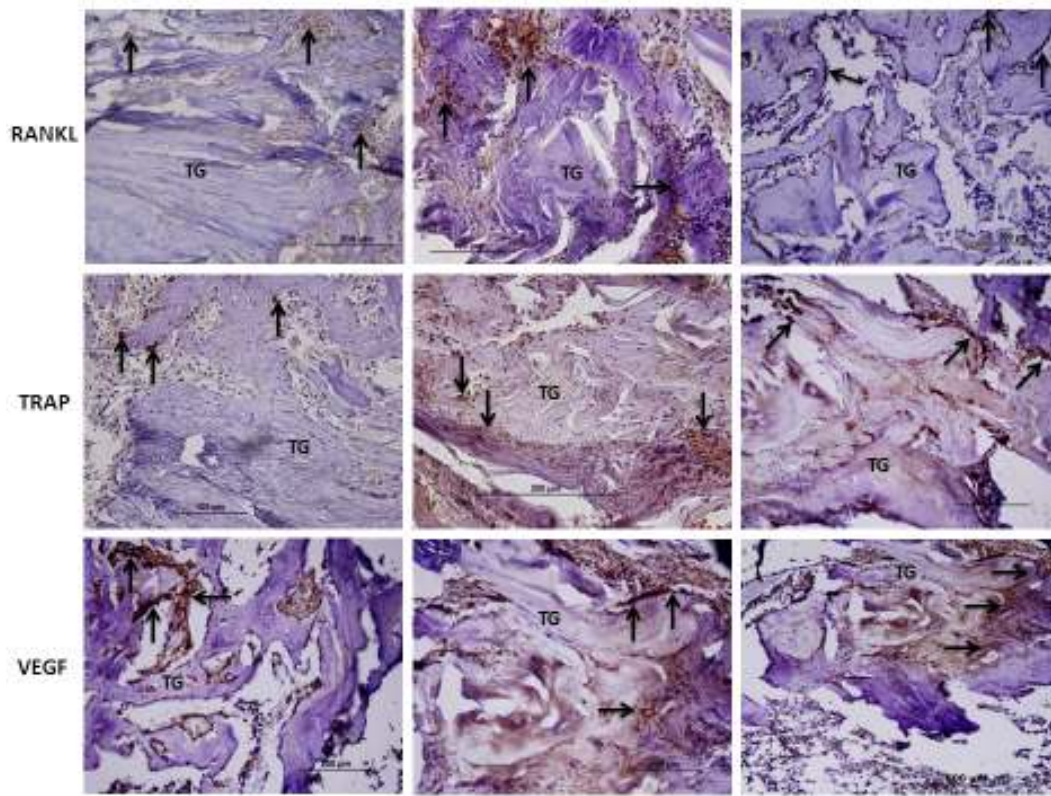
289 |



290 |

291 | **FIG. 3.** Immunolabeled proteins (arrows) used for assessing tendon graft (TG). ALP, alkaline  
292 | phosphatase; OPG, osteoprotegerin; OC, osteocalcin; ~~RANK, receptor activator of nuclear~~  
293 | ~~factor kB; TRAP, tartrate-resistant acid phosphatase; VEGF, vascular endothelial growth factor.~~  
294 | 7 days (left), 15 days (middle) and 30 days (right).





296

297

298 **Fig. 4.** Immunolabeled proteins (arrows) used for assessing tendon graft (TG). RANKL, receptor  
 299 activator of nuclear factor kB ligand; TRAP, tartrate-resistant acid phosphatase; VEGF, vascular  
 300 endothelial growth factor. 7 days (left), 15 days (middle) and 30 days (right).

301

302 4.

303

304 **4-DISCUSSION**

305

306 The present study aimed to analyze bone neoformation following new anionic bovine tendon  
 307 grafting in rat tibias, by means of morphological, histomorphometric and immunohistochemical  
 308 analyses. Anionic bovine tendon was shown to be an adequate and biocompatible material for  
 309 efficient bone regeneration, with osteogenic capabilities that allow it to be used as a scaffold for  
 310 bone repair.

311

312 | Studies investigating how to repair bone defects have found that scaffolds are frequently  
313 | needed to induce the growth of new bone tissue [14][44]. Researchers have been searching for  
314 | new, enhanced and increasingly more biocompatible materials to minimize complications in  
315 | bone repair.

316 | \_Biomaterials can present granulomatous inflammation on a chronic basis, which is intimately  
317 | connected with the healing process of bone implantation [16,27-29][46, 27-29]. Anionic collagen  
318 | displays high biocompatible and biodegradable potential, in addition to low antigenicity and low  
319 | levels of inflammatory reactions, thus enhancing bone neoformation [13,30] [43, 30].

320 | -

321 | After the native tendon is hydrolyzed, fibers are modified by opening the pores, favoring bone  
322 | cell migration and growth in the matrix, especially due to the generated anionic charge in  
323 | addition to the presence of growth factors [14, 31, 32]. Hydrolysis is also responsible for  
324 | removing cells that can cause dystrophic calcification, intense local inflammatory reactions and  
325 | foreign body reactions from the matrix [14,20,25,31-33][44, 20, 25, 33].

326 | \_The anionic tendon has low levels of inflammatory response which can be associated with  
327 | improvements in new bone formation, allowing the restoration of osseous defects may occur  
328 | within a shorter period such as, for example, within 15 days in this study, according Rocha et. al  
329 | (2002) that reported osteoblast deposition in its own matrix along the formed scaffold and  
330 | collagen removed since the remodeling, demonstrating the enhanced performance of anionic  
331 | collagen when compared to materials that must be reabsorbed in order to allow for bone  
332 | regeneration.

333 |

334 | Morphometric analysis aimed to verify bone and tissue neoformation [20,34,35] [20, 34, 35].

335 | The amount of neoformed bone tissue in the control group was significant at day 30. The  
336 | experimental group presented significant amounts at day 7 in comparison to day 30. This  
337 | finding was in accordance with that of a previous study [20] [20] that showed that new bone was  
338 | formed starting on day 7 and increased with the time after bone implantation. This study also  
339 | showed that formation of new bone over collagenous tendon fibers was evident from day 15  
340 | onwards. The results of the present study were also in accordance with those of Pan et al.  
341 | [36][36], who observed the presence of endochondral bone neoformation in all experimental

342 groups, and Uchida et al. [34][34], who found altered properties of bone composition, such as  
343 increased bone matrix formation, mineral concentration, cortical thickness and volume of  
344 trabecular bone.

345

346 Several studies have used immunohistochemistry to analyze bone neoformation [37-42][37-42].  
347 Osteoblasts express ALP, which plays a very important role in the mineralization of the bone  
348 matrix [42, 43]. In the present study, ALP was present during all of the analyzed periods, thus  
349 demonstrating constant bone mineralization, as was the case in other studies that demonstrated  
350 high mineralization levels at the end of experiments [42,43][42].

351

352 Vascular endothelial growth factor indicates that vascularization in the receptor bed is occurring  
353 at a constant rate [40][40]; it was observed in the present study as a marker in all the analyzed  
354 periods, reaching peak intensity at 15 and 30 days. Another study [40][40] found that VEGF  
355 reached its highest level at 10 days, but decreased after day 20 until reaching a statistically  
356 significant difference after 60 days in the experimental group. As explained by Carano and  
357 Filvaroff [44][44] and Hankenson et al. [45][45], angiogenesis is essential to bone regeneration,  
358 in that it provides cells, oxygen, nutrients and growth factors to the implantation site.  
359 Corroborating the findings of the present study, Miguel et al. [16][16] diverged from other  
360 authors and did not find any evidence of vascular formation at the points surrounding  
361 mineralization nuclei between matrix fibers.

362

363 The use of TRAP expressed by osteoclasts provides the rate of bone remodeling [41,46-48][41,  
364 46-48]. In the present findings, this marker was more prevalent in the experimental group on  
365 days 15 and 30. Pedrosa et al. [40][40] demonstrated similar TRAP curves in their control and  
366 experimental groups, with a maximum peak at 10 days. Furthermore, they noted that a constant  
367 level of receptor bed vascularization throughout the experiment showed that graft remodeling  
368 was occurring at a proportional rate.

369

370 The occurrence of osteoclastogenesis is demonstrated by the presence of RANKL and OPG  
371 [40,49,50][40, 49, 50], which in the present study were present at similar levels during all three

372 assessed periods; OPG presented peak intensity at the biomaterial-tissue interface at 7 days,  
373 and RANKL at 15 days. The present results were in agreement with previous research  
374 ~~[16,44,45]~~~~[16, 44, 45]~~ regarding bone regeneration, narrowing the gap between neoformation of  
375 the vascular bed and osteogenesis.

376

## 377 **5. CONCLUSION**

378 In conclusion,

379

### 380 **5- Conclusions**

381

382 the ~~The~~ findings of this study showed that anionic bovine tendon is an adequate and  
383 biocompatible material for efficient bone regeneration, with osteogenic capabilities that allow it  
384 to be used as a scaffold for bone repair.

385

## 386 **COMPETING INTERESTS**

387 Authors have declared that no competing interests exist.

388

## 389 **AUTHORS' CONTRIBUTIONS:**

390

391 This work was carried out in collaboration between all authors. Authors RLB, JCA, DVB, ACR,  
392 JSR, DDR and GG participated in concept and design, data collection and data  
393 analysis/interpretation. RO participated performing immunohistochemical analysis. MPOR, ALS  
394 and GMRJr participated in manuscript creation involving critical writing and revising of the  
395 content. All authors read and approved the final version of this manuscript.

396

## 397 **ETHICAL APPROVAL**

398

399 The study was approved by the ethics committee of the University of Marília (Marília, São  
400 Paulo, Brazil).

401

402

## 403 **↓-REFERENCES**

404

405  
406  
407  
408  
409  
410  
411  
412  
413  
414  
415  
416  
417  
418  
419  
420  
421  
422  
423  
424  
425  
426  
427  
428  
429  
430  
431  
432  
433

1. [Blum JS, Barry MA, Mikos AG. Bone regeneration through transplantation of genetically modified cells. Clin Plast Surg. 2003;30:611-20.](#)
2. [Liu J, Cao Z, Li C. Intermittent PTH administration: a novel therapy method for periodontitis-associated alveolar bone loss. Med Hypotheses. 2009;72:294-6.](#)
3. [Clarke BL, Khosla S. Physiology of bone loss. Radiol Clin North Am. 2010;48:483-95.](#)
4. [Iqbal J, Sun L, Zaidi M. Commentary-FSH and bone 2010: evolving evidence. Eur J Endocrinol. 2010;63:173-6.](#)
5. [Hardy R, Cooper MS. Bone loss in inflammatory disorders. J Endocrinol. 2009;201:309-20.](#)
6. [Petite H, Viateau V, Bensaïd W, Meunier A, de Pollak C, Bourguignon M, et al. Tissue-engineered bone regeneration. Nat Biotechnol. 2000;18:959-63.](#)
7. [Manolagas SC, Jilka RL. Bone marrow, cytokines, and bone remodeling. Emerging insights into the pathophysiology of osteoporosis. N Engl J Med. 1995;332:305-11.](#)
8. [Raggatt LJ, Partridge NC. Cellular and molecular mechanisms of bone remodeling. J Biol Chem. 2010;285:25103-8.](#)
9. [Schopper C, Moser D, Spassova E, Goriwoda W, Lagogiannis G, Hoering B, et al. Bone regeneration using a naturally grown HA/TCP carrier loaded with rh BMP-2 is independent of barrier-membrane effects. J Biomed Mater Res A. 2008;85:954-63.](#)
10. [Babensee JE, McIntire LV, Mikos AG. Growth factor delivery for tissue engineering. Pharm Res. 2000;17:497-504.](#)
11. [Gasque SCK, Correa AM, Cestari TM, Taga R, Oliveira RC, Zambuzzi WF, et al. Collagenic matrix from bovine tendon as a potential biomaterial for tissue bioengineering. Innov Implant J. Biomater Esthet. 2011;6:16-20.](#)
12. [Ruszczak Z, Friess W. Collagen as a carrier for on-site delivery of antibacterial drugs. Adv Drug Deliv Rev. 2003;55:1679-98.](#)
13. [Moreira PL, An YH, Santos AR Jr, Genari SC. In vitro analysis of anionic collagen scaffolds for bone repair. J Biomed Mater Res B Appl Biomater. 2004;71:229-37.](#)



- 434 14. Rocha LB, Goissis G, Rossi MA. Biocompatibility of anionic collagen matrix as scaffold  
435 for bone healing. Biomaterials. 2002;23:449-56
- 436 15. Martins VCA, Goissis G. Anionic collagen as matrix for oriented deposition of Calcium  
437 Phosphate Minerals. Polímeros: Ciência e Tecnologia. 1996;6:30-77.
- 438 16. Miguel FB, Barbosa Júnior Ade A, de Paula FL, Barreto IC, Goissis G, Rosa FP.  
439 Regeneration of critical bone defects with anionic collagen matrix as scaffolds. J Mater  
440 Sci Mater Med. 2013;24:2567-75.
- 441 17. Angele P, Abke J, Kujat R, Faltermeier H, Schumann D, Nerlich M, et al. Influence of  
442 different collagen species on physico-chemical properties of crosslinked collagen  
443 matrices. Biomaterials. 2004;25:2831-41.
- 444 18. Charulatha V, Rajaram A. Influence of different crosslinking treatments on the physical  
445 properties of collagen membranes. Biomaterials. 2003;24:759-67.
- 446 19. Goissis G, Giglioti A de F, Braile DM. Preparation and characterization of an acellular  
447 bovine pericardium intended for manufacture of valve bioprostheses. Artif Organs.  
448 2011;35:484-9.
- 449 20. Buchaim RL, Goissis G, Andreo JC, Roque DD, Roque JS, Buchaim DV, et al.  
450 Biocompatibility of anionic collagen matrices and its influence on the orientation of  
451 cellular growth. Cienc Odontol Bras. 2007;10:12-20.
- 452 21. Aparecida de Aro A, Vidal Bde C, Pimentel ER. Biochemical and anisotropical  
453 properties of tendons. Micron. 2012;43:205-14.
- 454 22. Quesada GAT, Brenner FB, Feltraco LT. Analysis of bovine collagen membrane,  
455 compared with expanded polytetrafluoroethylene membrane, used as protection  
456 barrierregenerations for further guided bone implant placement and treatment of  
457 periimplantitis with and without use of grafts. Rev Dentística on line. 2011;10:29-38.
- 458 23. Goissis G, Piccirilli L, Goes JC, de Guzzi Plepis AM, Das-Gupta DK. Anionic collagen:  
459 polymer composites with improved dielectric and rheological properties. Artif Organs.  
460 1998;22:203-9.
- 461 24. Lacerda C, Plepis AMG, Goissis G. Selective hydrolysis of carboxyamides of  
462 asparagine and glutamine residues of collagen: preparation and characterization of  
463 anionic collagen matrices for biomaterial applications. Quim Nova. 1998;21:267-71.

- 464 25. Bet MR, Goissis G, Lacerda CA. Characterization of polyanionic collagen prepared by  
465 selective hydrolysis of asparagine and glutamine carboxyamide side chains.  
466 Biomacromolecules. 2001;2:1074-9.
- 467 26. Mandarin-de-Lacerda Carlos A. Stereological tools in biomedical research. An. Acad.  
468 Bras. Ciênc. 2003;75:469-486.
- 469 27. Tsai AT, Rice J, Scatena M, Liaw L, Ratner BD, Giachelli CM. The role of osteopontin  
470 in foreign body giant cell formation. Biomaterials. 2005;26:5835-43.
- 471 28. Rosa FP, Lia RC, de Souza KO, Goissis G, Marcantonio E Jr. Tissue response to  
472 polyanionic collagen: elastin matrices implanted in rat calvaria. Biomaterials.  
473 2003;24:207-12.
- 474 29. Anderson JM, Rodriguez A, Chang DT. Foreign body reaction to biomaterials. Semin  
475 Immunol. 2008;20:86-100.
- 476 30. Cunha MR, Santos AR Jr, Goissis G, Genari SC. Implants of polyanionic collagen  
477 matrix in bone defects of ovariectomized rats. J Mater Sci Mater Med. 2008;19:1341-  
478 8.
- 479 31. Rocha LB, Brochi MA, Bellucci AD, Rossi MA. Efficacy of polyanionic collagen  
480 matrices for bone defect healing. J Biomed Mater Res B Appl Biomater. 2004;71:355-  
481 9.
- 482 32. Goissis G, da Silva Maginador SV, da Conceição Amaro Martins V. Biomimetic  
483 mineralization of charged collagen matrices: in vitro and in vivo study. Artif Organs.  
484 2003;27:437-43.
- 485 33. Bet MR, Goissis G, Vargas S, Selistre-de-Araujo HS. Cell adhesion and cytotoxicity  
486 studies over polyanionic collagen surfaces with variable negative charge and  
487 wettability. Biomaterials 2003;24:131-7.
- 488 34. Uchida R, Bhawal UK, Kiba H, Arai K, Tanimoto Y, Kuboyama N, Asakura et al. Effect  
489 of plasma-irradiated silk fibroin in bone regeneration. J Biosci Bioeng. 2014;118:333-  
490 40.
- 491 35. Zaker Shahrak A, Zor F, Kanatas A, Acikel C, Sapountzis S, Nicoli F, et al.  
492 Morphological and morphometric evaluation of the ilium, fibula, and scapula bones for  
493 oral and maxillofacial reconstruction. Microsurgery. 2014;34:638-45.

- 494 36. Pan W, Cao Z, Li D, Zhang M. Evaluation of the potential application of three different  
495 biomaterials combined with bone morphological proteins for enhancing tendon-bone  
496 integration. Injury. 2013;44:550-7.
- 497 37. Alberius P, Gordh M, Lindberg L, Johnell O. Onlay bone graft behaviour after marrow  
498 exposure of the recipient rat skull bone. Scand J Plast Reconstr Surg Hand Surg.  
499 1996;30:257-66.
- 500 38. Kleinheinz J, Stratmann U, Joos U, Wiesmann HP. VEGF-activated angiogenesis  
501 during bone regeneration. J Oral Maxillofac Surg. 2005;63:1310-6.
- 502 39. Faria PE, Okamoto R, Bonilha-Neto RM, Xavier SP, Santos AC, Salata LA.  
503 Immunohistochemical, tomographic and histological study on onlay iliac grafts  
504 remodeling. Clin Oral Implants Res. 2008;19:393-401.
- 505 40. Pedrosa WF Jr, Okamoto R, Faria PE, Arnez MF, Xavier SP, Salata LA.  
506 Immunohistochemical, tomographic and histological study on onlay bone graft  
507 remodeling. Part II: calvarial bone. Clin Oral Implants Res. 2009;20:1254-64.
- 508 41. Nagata MJ, de Campos N, Messoria MR, Santinoni CS, Bomfim SR, Fucini SE, et al.  
509 Platelet-Rich Plasma Derived From Bone Marrow Aspirate Promotes New Cementum  
510 Formation. J Periodontol. 2014;7:1-17.
- 511 42. Toda M, Ohno J, Shinozaki Y, Ozaki M, Fukushima T. Osteogenic potential for  
512 replacing cells in rat cranial defects implanted with a DNA/protamine complex paste.  
513 Bone. 2014;67:237-45.
- 514 43. Sugawara Y, Suzuki K, Koshikawa M, Ando M, Iida J. Necessity of enzymatic activity  
515 of alkaline phosphatase for mineralization of osteoblastic cells. Jpn J Pharmacol.  
516 2002;88:262-9.
- 517 44. Carano RA, Filvaroff EH. Angiogenesis and bone repair. Drug Discov Today.  
518 2003;8:980-9.
- 519 45. Hankenson KD, Dishowitz M, Gray C, Schenker M. Angiogenesis in bone  
520 regeneration. Injury. 2011;42:556-61.
- 521 46. Yaziji H, Janckila AJ, Lear SC, Martin AW, Yam LT. Immunohistochemical detection of  
522 tartrate-resistant acid phosphatase in non-hematopoietic human tissues. Am J Clin  
523 Pathol. 1995;104:397-402.

- 524 47. Janckila AJ, Neustadt DH, Yam LT. Significance of serum TRACP in rheumatoid  
525 arthritis. J Bone Miner Res. 2008;23:1287-95.
- 526 48. Cheng T, Wang M, Chen Z, Eisenberg RA, Zhang Y, Zou Y, et al. Tartrate-resistant  
527 acid phosphatase 5b is a potential biomarker for rheumatoid arthritis: a pilot study in  
528 Han Chinese. Chin Med J. (Engl) 2014;127:2894-9.
- 529 49. Saraiva GL, Lazaretti-Castro M. Biochemical Bone Markers in Clinical Practice. Arg  
530 Bras Endocrinol Metab. 2002;46:72-8.
- 531 50. Pereira A, Vaz P, Rocha G, Felino A, Tavares P. Genetic engineering in implantology  
532 - the rankl. Rev Port Estomatol Med Dent Cir Maxilofac. 2011;52:170-4.
- 533
- 534 ~~2. Blum JS, Barry MA, Mikos AG. Bone regeneration through transplantation of genetically~~  
535 ~~modified cells. Clin Plast Surg. 2003;30:611-20.~~
- 536 ~~3. Liu J, Cao Z, Li C. Intermittent PTH administration: a novel therapy method for periodontitis-~~  
537 ~~associated alveolar bone loss. Med Hypotheses. 2009;72:294-6.~~
- 538 ~~4. Clarke BL, Khosla S. Physiology of bone loss. Radiol Clin North Am. 2010;48:483-95.~~
- 539 ~~5. Iqbal J, Sun L, Zaidi M. Commentary-FSH and bone 2010: evolving evidence. Eur J~~  
540 ~~Endocrinol. 2010;63:173-6.~~
- 541 ~~6. Hardy R, Cooper MS. Bone loss in inflammatory disorders. J Endocrinol. 2009;201:309-20.~~
- 542 ~~7. Petite H, Viateau V, Bensaïd W, Meunier A, de Pollak C, Bourguignon M, et al. Tissue-~~  
543 ~~engineered bone regeneration. Nat Biotechnol. 2000;18:959-63.~~
- 544 ~~8. Manolagas SC, Jilka RL. Bone marrow, cytokines, and bone remodeling. Emerging insights~~  
545 ~~into the pathophysiology of osteoporosis. N Engl J Med. 1995;332:305-11.~~
- 546 ~~9. Raggatt LJ, Partridge NC. Cellular and molecular mechanisms of bone remodeling. J Biol~~  
547 ~~Chem. 2010;285:25103-8.~~
- 548 ~~10. Schopper C, Moser D, Spasova E, Goriwoda W, Lagogiannis G, Hoering B, et al. Bone~~  
549 ~~regeneration using a naturally grown HA/TCP carrier loaded with rh BMP-2 is independent of~~  
550 ~~barrier-membrane effects. J Biomed Mater Res A. 2008;85:954-63.~~

- 551 ~~11. Babensee JE, McIntire LV, Mikos AG. Growth factor delivery for tissue engineering. Pharm~~  
552 ~~Res. 2000;17:497-504.~~
- 553 ~~12. Gasque SCK, Correa AM, Cestari TM, Taga R, Oliveira RC, Zambuzzi WF, et al. Matriz~~  
554 ~~colagênica de tendão bovino como potencial biomaterial para bioengenharia de tecidos.~~  
555 ~~Innov Implant J. Biomater Esthet. 2011;6:16-20.~~
- 556 ~~13. Ruszczak Z, Friess W. Collagen as a carrier for on-site delivery of antibacterial drugs. Adv~~  
557 ~~Drug Deliv Rev. 2003;55:1679-98.~~
- 558 ~~14. Moreira PL, An YH, Santos AR Jr, Genari SC. In vitro analysis of anionic collagen scaffolds~~  
559 ~~for bone repair. J Biomed Mater Res B Appl Biomater. 2004;71:229-37.~~
- 560 ~~15. Rocha LB, Goissis G, Rossi MA. Biocompatibility of anionic collagen matrix as scaffold for~~  
561 ~~bone healing. Biomaterials. 2002;23:449-56~~
- 562 ~~16. Martins VCA, Goissis G. Colágeno Aniônico como Matriz para Deposição Orientada de~~  
563 ~~Minerais de Fosfato de Cálcio. Polímeros: Ciência e Tecnologia. 1996;6:30-77.~~
- 564 ~~17. Miguel FB, Barbosa Júnior Ade A, de Paula FL, Barreto IC, Goissis G, Rosa FP.~~  
565 ~~Regeneration of critical bone defects with anionic collagen matrix as scaffolds. J Mater Sci~~  
566 ~~Mater Med. 2013;24:2567-75.~~
- 567 ~~18. Angele P, Abke J, Kujat R, Faltermeyer H, Schumann D, Nerlich M, et al. Influence of~~  
568 ~~different collagen species on physico-chemical properties of crosslinked collagen matrices.~~  
569 ~~Biomaterials. 2004;25:2831-41.~~
- 570 ~~19. Charulatha V, Rajaram A. Influence of different crosslinking treatments on the physical~~  
571 ~~properties of collagen membranes. Biomaterials. 2003;24:759-67.~~
- 572 ~~20. Goissis G, Giglioti A de F, Braille DM. Preparation and characterization of an acellular bovine~~  
573 ~~pericardium intended for manufacture of valve bioprostheses. Artif Organs. 2011;35:484-9.~~
- 574 ~~21. Buchaim RL, Goissis G, Andreo JC, Roque DD, Roque JS, Buchaim DV, et al.~~  
575 ~~Biocompatibility of anionic collagen matrices and its influence on the orientation of cellular~~  
576 ~~growth. Cienc Odontol Bras. 2007;10:12-20.~~

- 577 ~~22. Aparecida de Aro A, Vidal Bde C, Pimentel ER. Biochemical and anisotropical properties of~~  
578 ~~tendons. Micron. 2012;43:205-14.~~
- 579 ~~23. Quesada GAT, Brenner FB, Feltraco LT. Análise das membranas de colágeno bovino,~~  
580 ~~comparativamente às membranas de politetrafluoretileno expandido, como barreira de~~  
581 ~~proteção em regenerações ósseas guiadas para posterior colocação de implantes e no~~  
582 ~~tratamento de periimplantes com e sem o uso de enxertos bovinos. Rev Dentística on line.~~  
583 ~~2011;10:29-38.~~
- 584 ~~24. Goissis G, Piccirilli L, Goes JC, de Guzzi Plepis AM, Das-Gupta DK. Anionic collagen:~~  
585 ~~polymer composites with improved dielectric and rheological properties. Artif Organs.~~  
586 ~~1998;22:203-9.~~
- 587 ~~25. Lacerda C, Plepis AMG, Goissis G. Hidrólise seletiva de carboxiamidas de resíduos de~~  
588 ~~asparagina e glutamina em colágeno: preparação e caracterização de matrizes aniônicas~~  
589 ~~para uso como biomateriais. Quim Nova. 1998;21:267-71.~~
- 590 ~~26. Bet MR, Goissis G, Lacerda CA. Characterization of polyanionic collagen prepared by~~  
591 ~~selective hydrolysis of asparagine and glutamine carboxamide side chains.~~  
592 ~~Biomacromolecules. 2001;2:1074-9.~~
- 593 ~~27. Mandarin de Lacerda Carlos A. Stereological tools in biomedical research. An. Acad. Bras.~~  
594 ~~Ciênc. 2003;75:469-486.~~
- 595 ~~28. Tsai AT, Rice J, Scatena M, Liaw L, Ratner BD, Giachelli CM. The role of osteopontin in~~  
596 ~~foreign body giant cell formation. Biomaterials. 2005;26:5835-43.~~
- 597 ~~29. Rosa FP, Lia RC, de Souza KO, Goissis G, Marcantonio E Jr. Tissue response to~~  
598 ~~polyanionic collagen: elastin matrices implanted in rat calvaria. Biomaterials. 2003;24:207-~~  
599 ~~42.~~
- 600 ~~30. Anderson JM, Rodriguez A, Chang DT. Foreign body reaction to biomaterials. Semin~~  
601 ~~Immunol. 2008;20:86-100.~~

- 602 ~~31. Cunha MR, Santos AR Jr, Goissis G, Genari SC. Implants of polyanionic collagen matrix in~~  
603 ~~bone defects of ovariectomized rats. J Mater Sci Mater Med. 2008;19:1341-8.~~
- 604 ~~32. Rocha LB, Brochi MA, Bellucci AD, Rossi MA. Efficacy of polyanionic collagen matrices for~~  
605 ~~bone defect healing. J Biomed Mater Res B Appl Biomater. 2004;71:355-9.~~
- 606 ~~33. Goissis G, da Silva Maginador SV, da Conceição Amaro Martins V. Biomimetic~~  
607 ~~mineralization of charged collagen matrices: in vitro and in vivo study. Artif Organs.~~  
608 ~~2003;27:437-43.~~
- 609 ~~34. Bet MR, Goissis G, Vargas S, Solistre-de-Araujo HS. Cell adhesion and cytotoxicity studies~~  
610 ~~over polyanionic collagen surfaces with variable negative charge and wettability.~~  
611 ~~Biomaterials 2003;24:131-7.~~
- 612 ~~35. Uchida R, Bhawal UK, Kiba H, Arai K, Tanimoto Y, Kuboyama N, Asakura et al. Effect of~~  
613 ~~plasma-irradiated silk fibroin in bone regeneration. J Biosci Bioeng. 2014;118:333-40.~~
- 614 ~~36. Zaker Shahrak A, Zor F, Kanatas A, Acikel C, Sapountzis S, Nicoli F, et al. Morphological~~  
615 ~~and morphometric evaluation of the ilium, fibula, and scapula bones for oral and maxillofacial~~  
616 ~~reconstruction. Microsurgery. 2014;34:638-45.~~
- 617 ~~37. Pan W, Cao Z, Li D, Zhang M. Evaluation of the potential application of three different~~  
618 ~~biomaterials combined with bone morphological proteins for enhancing tendon-bone~~  
619 ~~integration. Injury. 2013;44:550-7.~~
- 620 ~~38. Alberius P, Gordh M, Lindberg L, Johnell O. Onlay bone graft behaviour after marrow~~  
621 ~~exposure of the recipient rat skull bone. Scand J Plast Reconstr Surg Hand Surg.~~  
622 ~~1996;30:257-66.~~
- 623 ~~39. Kleinheinz J, Stratmann U, Joos U, Wiesmann HP. VEGF-activated angiogenesis during~~  
624 ~~bone regeneration. J Oral Maxillofac Surg. 2005;63:1310-6.~~
- 625 ~~40. Faria PE, Okamoto R, Bonilha Neto RM, Xavier SP, Santos AC, Salata LA.~~  
626 ~~Immunohistochemical, tomographic and histological study on onlay iliac grafts remodeling.~~  
627 ~~Clin Oral Implants Res. 2008;19:393-401.~~

- 628 41. Pedrosa WF Jr, Okamoto R, Faria PE, Arnez MF, Xavier SP, Salata LA.  
629 Immunohistochemical, tomographic and histological study on onlay bone graft remodeling.  
630 Part II: calvarial bone. Clin Oral Implants Res. 2009;20:1254-64.
- 631 42. Nagata MJ, de Campos N, Messori MR, Santinoni CS, Bomfim SR, Fucini SE, et al.  
632 Platelet-Rich Plasma Derived From Bone Marrow Aspirate Promotes New Cementum  
633 Formation. J Periodontol. 2014;7:1-17.
- 634 43. Toda M, Ohno J, Shinozaki Y, Ozaki M, Fukushima T. Osteogenic potential for replacing  
635 cells in rat cranial defects implanted with a DNA/protamine complex paste. Bone.  
636 2014;67:237-45.
- 637 44. Sugawara Y, Suzuki K, Koshikawa M, Ando M, Iida J. Necessity of enzymatic activity of  
638 alkaline phosphatase for mineralization of osteoblastic cells. Jpn J Pharmacol. 2002;88:262-  
639 9.
- 640 45. Carano RA, Filvaroff EH. Angiogenesis and bone repair. Drug Discov Today. 2003;8:980-9.
- 641 46. Hankenson KD, Dishowitz M, Gray C, Schenker M. Angiogenesis in bone regeneration.  
642 Injury. 2011;42:556-61.
- 643 47. Yaziji H, Janckila AJ, Lear SC, Martin AW, Yam LT. Immunohistochemical detection of  
644 tartrate-resistant acid phosphatase in non-hematopoietic human tissues. Am J Clin Pathol.  
645 1995;104:397-402.
- 646 48. Janckila AJ, Neustadt DH, Yam LT. Significance of serum TRACP in rheumatoid arthritis. J  
647 Bone Miner Res. 2008;23:1287-95.
- 648 49. Cheng T, Wang M, Chen Z, Eisenberg RA, Zhang Y, Zou Y, et al. Tartrate-resistant acid  
649 phosphatase 5b is a potential biomarker for rheumatoid arthritis: a pilot study in Han  
650 Chinese. Chin Med J. (Engl) 2014;127:2894-9.
- 651 50. Saraiva GL, Lazaretti-Castro M. Marcadores Bioquímicos da Remodelação Óssea na  
652 Prática Clínica. Arq Bras Endocrinol Metab. 2002;46:72-8.



653 | ~~51.Pereira A, Vaz P, Rocha G, Folino A, Tavares P. Engenharia genética em implantologia — o~~  
654 | ~~rankl. Rev Port Estomatol Med Dent Cir Maxilofac. 2011;52:170–4.~~  
655 |