Rhoifolin; A Potent Antiproliferative Effect On Cancer Cell Lines

Omayma A. Eldahshan*

Pharmacognosy Department, Faculty of Pharmacy, Ain shams University, Cairo, Egypt

ABSTRACT

Aims: To investigate the cytotoxic activity of rhoifolin against different cancer cell lines **Study Design:** isolation, identification and cytotoxic activity evaluation.

Place and Duration of Study: Faculty of Pharmacy, Ain Shams University and Al-Azhar University, between October, 2010 and January, 2011.

Methodology: Rhoifolin, Apigenin 7-O-β neohesperidoside was isolated in a copious amount from the leaves of *Chorisia crispiflora* (Bombaceae). Its identity was unambiguously confirmed via different spectroscopic methods (UV, HNMR, CNMR and HMBC) and Viability assay test was used to evaluate its cytotoxic activity.

Results: It exhibited potent anticancer activities, nearly similar to that of vinblastine, when evaluated against human epidermoid larynex (Hep 2) and human cervical (HeLa) carcinoma cell lines. Promising activities were also obtained against hepatocellular (Hep G2), colon (HCT-116) and fetal human lung fibroblast (MRC-5) carcinoma cell lines. A unique effect of rhoifolin was in having no cytotoxic activity against healthy normal cells (Vero cells) which indicates a high selectivity of the compound selected.

Conclusion: The findings of this study showed that rhoifolin could be used as an ideal anticancer agent. It discriminates between cancerous and non cancerous cell as it kills only the former one. So the side effects which may appear during chemotherapy could be overcome.

Keywords: Roifolin; Chorisia crispiflora; Bombaceae; spectroscopic methods; cytotoxic activity

1. INTRODUCTION

The introduction of active agents derived from nature into the cancer armamentarium has changed the natural history of many types of human cancer. Throughout medical history, novel plant-derived compounds were of great significance to cancer therapy. As examples of these compounds are vinblastine and vincristine; Catharanthus roseus family Apocynaceae (Kalidass et al., 2010). Currently, one of the major cancer treatments is chemotherapy. Most of the chemotherapeutic drugs such as vincristine, paclitaxel, and etoposide (ET) cannot discriminate between cancer and non-cancer cells. Many normal cells are also killed during the process of chemotherapy. This nonspecific cytotoxicity damages the patient's immune system and generates many side effects such as neutropenia, vomiting, hair loss, peripheral neurotoxicity, etc. (Perry et al., 1976 and Einzig et al., 1991).

* Tel.: 002 010 11 8419 51

E-mail address: omiahm@hotmail.com

Rhoifolin is apigenin 7-O- β neohesperidoside. It was reported that rhoifolin has lots of pharmacological actions. It exerts its anti-diabetic effect through enhanced adiponectin secretion, phosphorylation of insulin receptor- β , and GLUT4 translocation (Rao *et al.*, 2011). It has an antiinflammatory action via multi-level regulation of inflammatory mediators (Eldahshan and Azab, 2012). Rhoifolin produced no change in hypoxic pulmonary vasoconstriction, but decreased cardiac output and aortic pressure (Occhiuto and Limardi, 1994).

Apigenin nucleus is a cancer chemopreventive agent. It inhibits cell proliferation in cancer cell types (Sarkar and Li, 2004). Because of its potential antioxidant, anti-inflammatory, and anti-tumor properties, apigenin is considered as a candidate cancer chemopreventive agent (Birt *et al.*, 1996; Birt *et al.*, 1997; Lepley *et al.*, 1996; Ross and Kasum, 2002).

Apigenin inhibited the growth through an apoptotic pathway in human cervical carcinoma HeLa cells (Zheng et al., 2005). Furthermore, apigenin inhibited A549 lung cancer cell proliferation and vascular endothelial growth factor (VEGF) transcriptional activation in a dose-dependent manner (Ling et al., 2005). However, Kawaii (Kawaii et al., 1999) reported the weak effect of rhoifolin against melanin pigment producing mouse melanoma, human T-cell leukemia, human lung and lymph node metastatic carcinoma cell lines.

So far, nothing has been documented about the cytotoxic effect of rhoifolin against human epidermoid larynex (Hep 2), human cervical (HeLa), hepatocellular (Hep G2), colon (HCT-116) and fetal human lung fibroblast (MRC-5) carcinoma cell lines, so the antineoplastic activity of this compound was investigated against these types of cancer cell lines. We reported here for the first time the high potent and selective antitumor activity of rhoifolin against several types of cancer cell lines to develop a preliminary building block for the construction of a new anticancer drug.

2. MATERIALS AND METHODS

2.1. Plant Material

- Chorisia leaves were collected from Zoo Garden in Giza, Egypt, 2010 and were authenticated by Prof. Dr Abdel Salam El Noyehy, Prof. of Taxonomy, Faculty of Science, Ain Shams University, Cairo, Egypt. Voucher specimen was deposited in the herbarium of Pharmacognosy Department (voucher specimen number; CCB-73), Faculty of Pharmacy, Ain Shams University, Cairo, Egypt.
- The leaves were dried in shade and milled to a fine powder.

,

2.2. Extraction and Isolation

- Powder of air dried leaves of *Chorisia crispiflora* (1 kg) was extracted with 70 % ethanol at room temperature. The extract was entirely dried and dissolved in a small amount of water and
- 69 partioned with *n*-hexane, ethyl acetate and butanol successively. The aqueous water residue
- 70 was totally dried and extracted with methanol at 40°C. The methanolic extract upon
- 71 concentration yielded yellow crystals of rhoifolin (8.3 g). Purification to the crystals was achieved
- 72 by crystallization.

2.3. Instruments and Materials for Phytochemical Investigation

- 74 Chromatographically pure materials 1 mg each were dissolved in analytically pure methanol next
- 75 subjected to UV spectroscopic investigation in 4 ml capacity quartz cells 1 cm thick using a Carl
- Zeiss spectrophotometer PMQ II. AlCl₃, AlCl₃/HCl, fused NaOAc / H₃BO₃ and NaOMe reagents
- 77 were separately added to the methanolic solution of investigated material and UV measurements
- 78 were later carried out.

- 79 The NMR spectra were recorded on a Varian Mercury VX-500 NMR spectrometer. H- spectra
- 80 ran at 300 MHz and ¹³C- spectra were run at 75.46 MHz in deutrated dimethylsulphoxide (DMSO-
- 81 d_6).
- 82 Rhoifolin: Apigenin 7-O-β neohesperidoside, C₂₇H₃₀O₁₄, yellow needles, m.p. 250-265°C.
- 83 IR vmax (KBr): 3388 (OH), 1657 (α, β-unsat. CO), 1605, 1497, and 1488 (arom. C=C), 1249,
- 84 1178, 1074 (glycosidic C–O) cm⁻¹.
- 85 UV λmax (log ε) (MeOH): 266 (4.20), 336 (4.30) nm; (MeONa): 267 (4.20), 387 (4.40) nm;
- 86 (NaOAc): 257 (4.20), 266 (4.20), 391 (4.40) nm, (NaOAc + H₃BO₃): 268 (4.20), 340 (4.30) nm;
- 87 (AlCl3): 275 (4.20), 299 (4.10), 350 (4.20), 385 (4.20) nm, (AlCl₃ + HCl): 276 (4.20), 298 (4.10),
- 88 342 (4.20), 382 (4.10) nm.
- 89 ¹H-NMR, DMSO- d_6 δ ppm: 7.91(2H, d, J=8.8 Hz, H-2`,6`), 6.92(2H, d, J= 8.8 Hz, H-3`,5`), 6.84
- 90 (1H, d, J= 2.0 Hz, H-8), 6.80 (1H, s, H-3), 6.33(1H, d, J= 2.0 Hz, H-6), 5.08(1H, singlet like, H-
- 91 1```), 5.20 (1H, d, *J*= 7.3 Hz, H-1``), 1.16(3H, d, *J*=6.3Hz, H3-6```).
- 92 ¹³C-NMR, DMSO-*d*₆ δ ppm: 182.1-C4, 164.4-C2, 162.6-C7, 161.7-C4`, 161.1-C5, 157.1-C9,
- 93 128.7-C2`,6`, 120.9-C1`, 116.2C-3`,5`, 105.5-C10, 103.2-C3, 99.4-C6, 94.6-C8 , Sugar proton:
- 94 100.5-C1``, 98.2-C1```, 77.6-C2``, 77.4-C3``, 76.8-C5``, 72.3-C4```, 71.0-C2```, 70.8-C3```, 70.1-
- 95 C4``, 68.8-C5```, 60.9-C-6``, 18.5-C-CH₃.

96 2.4. Mammalian Cell Lines

- 97 Vero cells (Normal kidney cells).
- 98 Carcinoma cell lines: Hep2 (human epidermoid larynex carcinoma cells), HeLa cells (human
- 99 cervical carcinoma cells). Hep G2 (human hepatocellular carcinoma). HCT-116 (human colon
- 100 carcinoma cells) and MRC-5 (fetal human lung fibroblast cells).
- 101 All cell lines of a well-differentiated carcinoma were obtained from the American Type Culture
- 102 Collection (ATCC).

107

111

113

103 2.5. Chemical Used

- 104 Dimethyl sulfoxide (DMSO), crystal violet and trypan blue dye (Sigma, St.Louis, Mo., USA).
- 105 DMEM, RPMI-1640, FBS, HEPES buffer solution, L-glutamine, gentamycin and 0.25 % Trypsin-
- 106 EDTA (Bio Whittaker ®Lonza, Belgium). Crystal violet stain (1%).

108 2.6. Cytotoxicity Evaluation Using Viability Assay

- 109 Cell toxicity was monitored by determining the effect of the test sample on cell viability through
- the viability test (Vijaya et al., 2004 and Mosmann, 1983).

112 3. RESULTS AND DISCUSSION

114 3.1. Identification of the Compound

- 115 Pure material of rhoifolin was obtained as an amorphous light yellow powder, which appeared as
- a dark purple spot on Paper chromatography (PC) and turned yellow upon exposure to ammonia

vapors, under short UV light (254 nm). Confirmation of the compound was achieved through UV shift reagents, ¹HNMR, ¹³CNMR and HMBC correlation (Fig. 1).

3.2. Cytotoxic Activity of the Compound

The tested compound showed marked toxic effects to the cancerous cell lines. It exerted cytotoxic activity to Hep 2 and HeLa cell lines at IC₅₀ 5.90 and 6.2 μ g/mL respectively (Fig. 2 B & C). HepG2 is affected but to a lesser extent by the compound at IC₅₀ 22.6 μ g/mL (Fig. 2-D). The least potent activities were to μ GC-116 and MRC-5 at IC₅₀ 34.8 and 44.6 μ g/mL respectively (Fig. 2 E & F).

Historically natural products have been an important source of antineoplastic drugs. Sixty percent of currently used antitumor agents are of natural origin, derived from plants, marine organisms and is a useful tool for the discovery of new potential anticancer agents from natural products. One of the important criteria for a therapeutic drug for cancer is to have minimum or no side effects on normal body cells of patients undergoing chemotherapy. This invariably implies that the drug should not only have high potent activity at lower concentrations but also should exhibit a high degree of selectivity.

Thus, development of novel selective drugs is an important and challenging task, and understanding the biological differences between normal and cancer cells is essential for achieving this goal.

The present *in vitro* study showed the ability of rhoifolin to exhibit a high degree cytotoxic activity to cancerous cells with great selectivity, where, as it is clear, that the compound has no cytotoxic activity against mammalian normal cells (Table 1, Fig 2A).

Table 1 showed that the compound of interest exhibited high cytotoxic activity to laryngeal cancer cells, at a very low IC_{50} = 5.9 μ g/mL followed by cervical at IC_{50} = 6.2 μ g/mL, both are nearly similar to that of vinblastine. Hepatic cell line was also affected but at a lesser extent by the toxicity of rhoifolin at IC_{50} 22.6 μ g/mL. The colon and the fetal human lung fibroblast cell lines are affected at IC_{50} 34.8 and 44.6 μ g/mL respectively.

The selectivity index (SI) was defined as the ratio of the IC₅₀ obtained from the experiment on normal cells vs. cancer cells. High selectivity was achieved when the SI was \geq 3 (Prayong *et al.*, 2008). As the Selective index (SI) demonstrates the differential activity of a pure compound, the greater the SI value is, the more selective it is. An SI value less than 2 indicates general toxicity of the pure compound (Koch *et al.*, 2005).

Based on this, the SI data shown in Table 2 indicates that rhoifolin exhibits a very high degree of cytotoxic selectivity at SI greater than 8.47 for laryngeal cell lines, followed by 8.06 in cervical and 2.21 in hepatic carcinoma cell lines. The other two carcinoma cell lines; colon and fetal human lung fibroblast are of little SI.

Fig. 1. HMBC correlations of Apigenin 7-*O*-β neohesperidoside (Rhoifolin)

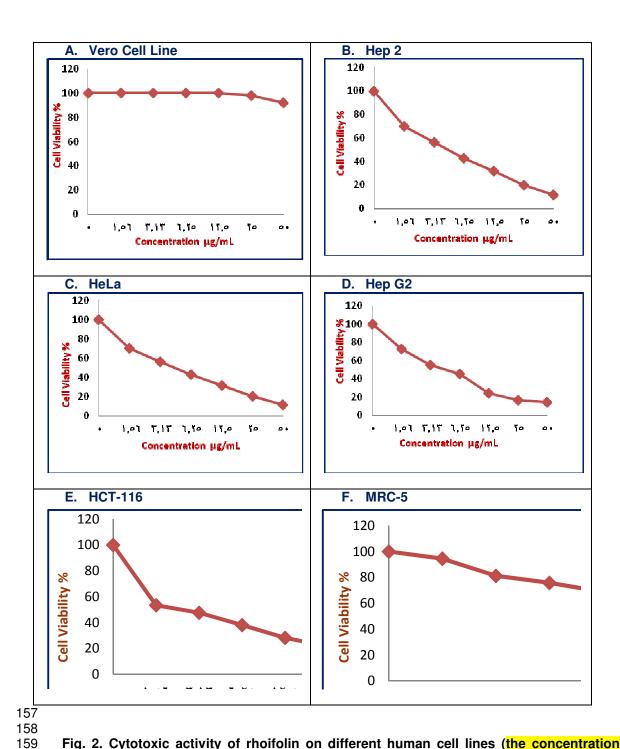


Fig. 2. Cytotoxic activity of rhoifolin on different human cell lines (the concentration in µg/mL): (A) Mammalian vero cell line (normal cell), (B) Hep2 (human epidermoid larynex carcinoma cells), (C) HeLa cells (human cervical carcinoma cells), (D) Hep G2 (human hepatocellular carcinoma), (E) HCT-116 (human colon carcinoma cells) and (F) MRC-5 (fetal human lung fibroblast cells).

Table 1. IC_{50} of rhoifolin and vinblastine on the carcinoma cell lines.

Cell	Rhoifolin Concentration		Vinl	olastine
	μg/mL	μМ	<mark>μg/mL</mark>	μМ
MRC-5	44.6	0.0770	4.6	0.0055
HCT	34.8	0.0601	2.6	0.0031
HepG2	22.6	0.0390	4.6	0.0055
HeLa	6.20	0.0107	5.2	0.0063
Hep2	5.90	0.0101	4.6	0.0055

Table 2. Selective indices of rhoifolin on carcinoma cell lines.

Cell lines	SI 173
	174
	175
MRC-5	> 1.12176
	177
	178
HCT	> 1.43179
	180
	181
HepG2	> 2.21 182
	183
	184
HeLa	> 8.06185
	186
	187
Hep2	> 8.47 188
op2	189
	190
	191

4. CONCLUSION

198 199

- 200 Interestingly, this present study, showed the following advantages of rhoifolin:
- 201 1. Potent cytotoxic effect nearly similar to that of vinblastine which may become a good 202 therapeutic strategy to its use as an antagonist for treatment of this dreaded disease, especially
- 203 laryngeal, cervical and hepatic cancer.
- 204 2. It is considered as an ideal antitumor agent to specific cancerous cells where it is toxic to
- 205 malignant with no toxicity to normal cells so it will be a good building unit for a new antitumor drug
- 206 without side effects.
- 207 However, currently there are limited numbers of such agents available for clinical use. The
- 208 mechanisms behind its respective anticancer effect are now under investigation to pave a way to
- 209 a discovery of a new cancer therapeutic agent.

210 211

ACKNOWLEDGEMENTS

212 213

The author is grateful for The Regional Center for Mycology and Biotechnology, Al-Azhar University, Cairo, Egypt for hosting the cytotoxic activities.

214 215 216

COMPETING INTERESTS

217 218

Author has declared that no competing interests exist.

219 220

REFERENCES

221 222

223 Birt, D.F., Pelling, J.C., Nair, S., Lepley, D. (1996). Diet intervention for modifying cancer risk. Prog. Clin. Biol. Res., 395, 223-234.

224

225

Birt, D.F., Mitchell, D., Gold, B., Pour, P., Pinch, H.C. 1997. Inhibition of ultraviolet light induced 226 227 skin carcinogenesis in SKH-1 mice by apigenin, a plant flavonoid. Anticancer Res., 17, 85-91.

228

- 229 Einzig, A.I., Hochster, H., Wiernik, P.H., Trump, D.L., Dutcher, J.P., Garowski, E. (1991). A 230 phase II study of taxol in patients with malignant melanoma. Invest New Drugs, 9, 59-64.
- 231 Eldahshan, O.A. and Azab, S.S. (2012). Anti-inflammatory Effect of Apigenin-7-
- 232 neohesperidoside (Rhoifolin) in Carrageenin- Induced Rat Oedema Model
- 233 Journal of Applied Pharmaceutical Science, 02 (08), 74-79.

234

- 235 Kalidass, C., Mohan, V.R., Daniel, A. (2010). Effect of auxin and cytokinin on vincristine
- 236 production by callus cultures of Catharanthus roseus L. (apocynaceae). Trop. Subtrop.
- 237 Agroecosystems, 12, 283-288.

- 239 Kawaii, S., Tomono, Y, Katase, E., Ogawa, K., Yano, M. (1999). Antiproliferative activity of
- 240 flavonoids on several cancer cell lines. Biosci Biotechnol Biochem., 63(5), 896-9.

Koch, A., Tamez, P., Pezzuto, J., Soejarto, D. (2005). Evaluation of plants used for antimalarial treatment by the Massai of Kenya. J Ethnopharmacol., 101, 95-99.

243

Lepley, D.M., Li, B., Birt, D.F., Pelling, J.C. (1996). The chemopreventive flavonoid apigenin induces G2/M arrest in keratinocytes. Carcinogenesis, 17, 2367–2375.

246

Liu, L.Z., Fang, J., Zhou, Q., Hu, X., Shi, X., Jiang, B.H. (2005). Apigenin inhibits expression of vascular endothelial growth factor and angiogenesis in human lung cancer cells: implication of chemoprevention of lung cancer. Mol Pharmacol., 68(3),635-43.

250

- Mosmann, T. (1993). Rapid colorimetric assay for cellular growth and survival:application to proliferation and cytotoxicity assays. J Immunol. Methods, 65, 55-63.
- Occhiuto, F. and Limardi, F. (1994). Comparative effects of the flavonoids luteolin, apiin and rhoifolin on experimental pulmonary hypertension in the dog. Phytotherapy Research, 8 (3), 153–156.

256

Perry, M.C., Moertel, C.G., Schutt, A.J., Reitemeier, R.J., Hahn, R.G. (1976). Phase II studies of dianhydrogalactitol and VP-16-213 in colorectal cancer. Cancer Treat Rep., 60, 1247–1250.

259

- Prayong, P., Barusrux, S., Weerapreeyakul, N. (2008). Cytotoxic activity screening of some indigenous Thai plants. Fitoterapia, 79, 598-601.
- Rao, Y.K., Lee, M., Chen, K., Lee, Y., Wu, W., Tzeng, W. (2009). Mimetic Action of Rhoifolin and
- Cosmosiin Isolated from *Citrus grandis* (L.) Osbeck Leaves: Enhanced Adiponectin Secretion and Insulin Receptor Phosphorylation in 3T3-L1 Cells. Evidence-Based Complementary and
- 265 Alternative Medicine Volume, 2011, 9 pages.

266 267

Ross, J.A., Kasum, C.M. (2002). Dietary flavonoids: bioavailability, metabolic effects, and safety. Annu. Rev. Nutr., 22, 19–34.

270

- Sarkar, F.H., Li, Y. (2004). Cell signaling pathways altered by natural chemopreventive agents. Mutat. Res., 555, 53–64.
- Vijaya, P., Raghu, C., Ashok, G., Dhanaraj, S.A. and Suresh, B. (2004). Antiviral activity of medicinal plants of Nilgiris. Indian J Med. Res, 120, 24-29.

275

Zheng, P.W., Chiang, L.C., Lin C.C. (2005). Apigenin induced apoptosis through p53-dependent pathway in human cervical carcinoma cells. Life Sci., 76, 1367–1379.